

MYCOBACTERIUM
CULTURE

Orderable - MYCCUL

Turnaround Time: 8 weeks (Microscopic results reported as soon as possible, culture requires up to 8 weeks)

Positives reported upon detection.

Alternate Name(s):

Acid Fast Bacilli Culture (AFB)

TB Culture

Specimen Type and Collection Instructions:



Laboratory:
Microbiology



Requisition:
See electronic order



Method of Analysis:
Microscopy, Culture, PCR

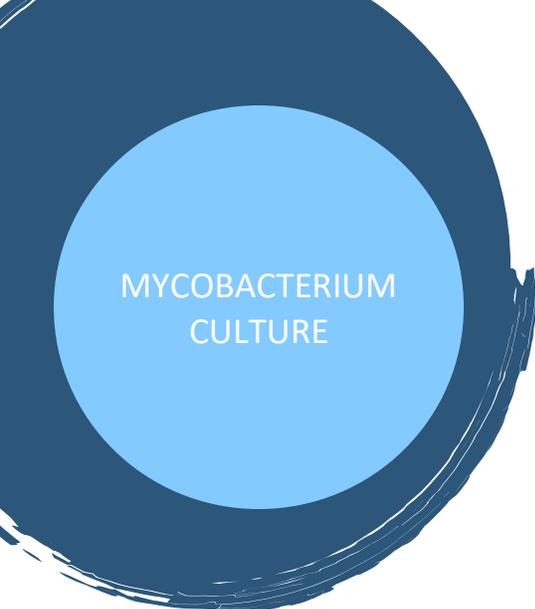


Test Schedule:
Referred out weekdays



Referred to:
Public Health Laboratory
1200 Commissioners Rd E
London, Ontario
519-455-9310

| Specimen Type | Collection instructions |
|--|--|
| Abscess contents, aspirated fluid | As much as possible (≥ 5 mL) in a sterile container |
| Blood | Collect into a 6 mL Dark Green (Sodium Heparin) Vacutainer tube (DO NOT SPIN) |
| Body fluids (pleural, pericardial, peritoneal, etc.) | Collect as much as possible (10-15 mL) in a sterile container |
| Bone | Collect in a sterile container |
| Bone Marrow | Collect into a 6 mL green top Vacutainer tube or sterile Container |
| Bronchial Alveolar Lavage (BAL) | Collect a minimum of 5 mL in a sterile container. Submit a post-bronchoscopy sputum as well |
| CSF | Collect a minimum of 2 mL in a sterile container. Specimens <0.5 mL will be rejected. |
| Faeces | 1.0 grams in sterile container |
| Gastric Lavage Fluid | Obtain Gastric Lavage transport media from the Microbiology Laboratory. Collect fasting, early morning sample to recover sputum swallowed during sleep. Following collection, the sample must be neutralized immediately by transferring it to the transport container which contains sodium carbonate. |



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| Specimen Type | Collection instructions |
|--------------------------------------|--|
| Induced Sputum | Collect 5-10 mL of induced sputum in a sterile container. Use hypertonic saline. Patient can expectorate several times per collection to achieve minimum volume. Ensure patient does NOT rinse mouth with tap water before expectoration or expectorate saliva or postnasal discharge. Avoid sputum contamination with nebulizer reservoir water. Do not pool samples. |
| Sputum | Collect 5-10 mL of expectorated or induced sputum in a sterile container. Patient can expectorate several times per collection to achieve minimum volume. Ensure patient does NOT rinse mouth tap water before expectoration or expectorate saliva or postnasal discharge. Do not pool samples. |
| Tissue Biopsy Sample | Collect 1 gram of tissue in a sterile container. Select caseous portion if available. A small amount (<1 mL) of sterile saline may be added to keep specimen from drying. Tissue submitted in anaerobic transport is not acceptable for culture. |
| Tracheal aspirate | Collect as much as possible into a sterile container. |
| Urine | Send minimum 40mL of first morning urine (midstream or catheter collection) in a sterile container. |

For more specimen types and detailed collection information, please see [Additional Mycobacterium Collection Information](#) from the Public Health Laboratory website.

Storage and Shipment:

Do not refrigerate blood for mycobacterial culture, hold at room temperature.

Refrigerate all other specimens whenever possible if transport to the laboratory or specimen processing will be delayed more than one hour.

The specimen must be received within 24 hours of collection.